



Cognitive Vitality Reports® are reports written by neuroscientists at the Alzheimer's Drug Discovery Foundation (ADDF). These scientific reports include analysis of drugs, drugs-in-development, drug targets, supplements, nutraceuticals, food/drink, non-pharmacologic interventions, and risk factors. Neuroscientists evaluate the potential benefit (or harm) for brain health, as well as for age-related health concerns that can affect brain health (e.g., cardiovascular diseases, cancers, diabetes/metabolic syndrome). In addition, these reports include evaluation of safety data, from clinical trials if available, and from preclinical models.

Trichostatin-A

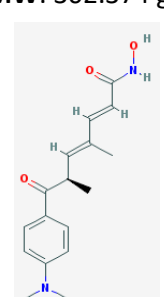
Evidence Summary

Broad-spectrum HDAC inhibitor that can act as a short-term cognitive enhancer. May extend lifespan and protect against acute oxidative stress damage. First human Phase 1 trial for cancer is in progress.

Neuroprotective Benefit: Can induce temporary cognitive boosting effects in animals by modulating expression of synaptic plasticity genes, but does not alleviate disease pathology or neuron loss.

Aging and related health concerns: Can extend lifespan in flies and worms. May be beneficial for cancer in combination therapy, and protective against oxidative stress damage when administered acutely after ischemic injury.

Safety: Limited information about long-term safety in animals, and initial safety testing in humans is ongoing. Has pleiotropic effects, which may include inhibition of oligodendrocyte differentiation and altered expression of pro-atherosclerotic genes.

Availability: Research use	Dose: Not established in humans	Chemical formula: C ₁₇ H ₂₂ N ₂ O ₃ MW: 302.374 g/mol  Source: Pubchem
Half-life: 10 minutes (plasma, in mice)	BBB: low penetrance	
Clinical trials: None	Observational studies: None	

What is it? Trichostatin-A is an antifungal antibiotic isolated from *Streptomyces hydgroscopius*. It acts as a potent broad-spectrum reversible inhibitor of class I and class II histone deacetylase enzymes [1]. It also acts as a cell cycle inhibitor. It has not been used in humans, but animal studies provide evidence for neuroprotection, lifespan extension, and anti-tumor activity. It mediates these effects by altering the epigenetic regulation of genes involved in these processes.

Neuroprotective Benefit: Can induce temporary cognitive boosting effects in animals by modulating expression of synaptic plasticity genes, but does not alleviate disease pathology or neuron loss.

Types of evidence:

- Numerous laboratory studies

Human research to suggest prevention of dementia, prevention of decline, or improved cognitive function? None

Human research to suggest benefits to patients with dementia: None

Mechanisms of action for neuroprotection identified from laboratory and clinical research:

Neurodegenerative diseases: Potential benefit (rodents)

As a histone deacetylase (HDAC) inhibitor, trichostatin-A, can affect the regulation and expression of genes involved in synaptic function and **act as a temporary cognitive enhancer**. In many



neurogenerative diseases, gene expression is altered due to an imbalance of histone acetylation, and trichostatin-A can help prevent or reverse these epigenetic genes and restore expression of genes important for the maintenance of cognitive function. The cognitive enhancing effects appear to stem primarily from trichostatin-A induced increases in CREB-associated genes, which are critical for regulating synaptic plasticity, and increased expression of brain derived neurotrophic factor (BDNF). However, the **effects of trichostatin-A are pleiotropic**, and only some of these changes are neuroprotective. The specific effects in a given cell population will depend on their epigenetic status, cell type, and tissue-related environmental factors. While trichostatin-A can temporarily improve performance on memory tasks, it **cannot promote regeneration or recover neuronal/synaptic loss** [2; 3].

Alzheimer's disease: Significant overlap was found in the genes regulated by trichostatin-A and those dysregulated in Alzheimer's disease (AD), in a microarray study [4]. Trichostatin-A treatment has shown to have some beneficial effects in rodent AD models by increasing expression of BDNF and rescuing hippocampal synaptic plasticity [5; 6]. However, these **benefits are generally short-lived and not associated with reductions in AD-associated pathology** or recovery of neuronal loss.

In rats injected with amyloid fibrils, hippocampal injection of trichostatin-A three days before testing led to improvements in escape latency on the Morris water maze and preference on the novel object recognition tasks [5]. This was accompanied by a recovery of BDNF expression driven by restoration of histone H3 acetylation at the BDNF promoter. In the APP/PS1 transgenic AD mouse model, acute trichostatin-A administration two hours prior to testing was able to restore hippocampal histone H4 acetylation levels, which are typically reduced by about 50% in these mice, improve performance on a contextual freezing task, and restore synaptic plasticity (Long-term potentiation) [6]. Chronic trichostatin-A administration (i.p. 5 mg/kg every other day for 2 months) promoted brain expression of gesolin, which has anti-amylogenic properties. However, due the pleiotropic effects of trichostatin-A, including an increase in secretase activity, it did not change the overall amyloid load in the brains of these animals [7].

Parkinson's disease: The neuroprotective potential of trichostatin-A depends on the nature of the neuronal insult, and in some contexts may instead exacerbate damage.

Pre-treatment with trichostatin-A, one-hour prior to MPTP, protected against nigrostriatal dopaminergic pathway neurodegeneration. Trichostatin-A relieved the repression on the neuron-restrictive silencer factor (NRSF) target genes, leading to an upregulation of BDNF and thyroid hormone [8]. Thyroid hormone (T3) suppresses expression of APP [9], which helps prevent A β production. In cultured SH-SY5Y dopaminergic-like cells, trichostatin-A could protect against MPP+-mediated mitochondrial



fragmentation by preventing the downregulation of Mfn2, but was not protective against rotenone-induced mitochondrial fragmentation [10]. Furthermore, trichostatin-A was found to exacerbate rotenone-mediated neurodegeneration in other dopaminergic cell lines [11].

Huntington's disease: In a transgenic Huntington's disease mouse model (HdhQ7/Q111), acute administration of trichostatin-A improved performance on a novel object recognition task (from $53 \pm 3\%$ to $60.5 \pm 3.5\%$) and rescued expression of CREB target genes in the hippocampus [12]. Trichostatin-A also increased vesicular transport of BDNF in striatal neurons via its inhibition of HDAC6 [13].

Stroke: Potential benefit (rodents)

Trichostatin-A is protective in the context of ischemic damage through its ability to promote **induction of the Nrf2 antioxidant system** through P13K/Akt signaling.

In the MCAO stroke model, trichostatin-A pre-treatment reduced infarct volume (from 49.1 ± 3.8 to $21.3 \pm 4.6\%$), edema, and neurological deficit scores [14; 15]. Since the mechanism appears to involve the reduction of oxidative stress damage, it seems likely that trichostatin-A would have to be administered very close to time of damage for it to be effective.

APOE4 interactions:

Trichostatin-A may be beneficial in alleviating ApoE4-associated endosomal dysfunction. Excessive endocytic acidification can promote APP processing and inhibit A β clearance [16]. Alkalinization of the endosomal compartment can attenuate APP processing and A β secretion. The endosomes in ApoE4 astrocytes are too acidic due to the downregulation of the Na⁺/H⁺ exchanger NHE6, possibly mediated by elevated HDAC4 activity [17]. Epigenetic modification of NHE6 restores its expression, which leads to alkalinization of the endosome and restoration of Lrp1 surface expression. Lrp1 is a receptor crucial to the phagocytic activity of astrocytes and helps mediate A β clearance.

Aging and related health concerns: Can extend lifespan in flies and worms. May be beneficial for cancer in combination therapy, and protective against oxidative stress damage when administered acutely after ischemic injury.

Types of evidence:

- Numerous laboratory studies



Lifespan: Benefit (flies and worms)

Trichostatin-A has been demonstrated to extend lifespan in *Drosophila* and *C. elegans* in a calorie restriction-like manner.

Trichostatin-A was shown to extend the lifespan of wild-type *C. elegans* by 22.12% (from 23.1 to 28.4 days), but did not further extend lifespan of long-lived eat-2 mutant worms, suggesting that trichostatin-A promotes lifespan through a similar mechanism to calorie restriction [18]. In *Drosophila*, trichostatin-A extended the maximum survival of male flies by 37.0% and female flies by 37.9% [19]. Notably, trichostatin-A relieved repression on the Hsp22 and Hsp70 promoters, both of which are more highly expressed in naturally long-lived flies than in short-lived fly strains [20]. This suggests that the optimal dose and effects are likely to vary based on genetic background.

The lifespan extension may involve the same induction of NHE6 mediated vacuolar alkalization shown to be beneficial in promoting A β clearance [17]. Trichostatin-A inhibits the HDAC Rpd3, leading to an increase in NHE6, which is a CREB-target gene that regulates cell responses under low-nutrient conditions [21].

Cancer: Potential benefit (rodents, cell culture)

Trichostatin-A has anti-tumorigenic properties due to its ability **to inhibit the cell cycle** and affect the expression of genes dysregulated in cancer. It may be most beneficial by working in a synergistic manner with other therapies to augment their anti-tumor responses.

Based on microarray analysis, the pool of genes regulated by trichostatin-a is largely comprised of cell-cycle and cancer-associated genes [4]. Due to differences in their epigenetic and expression profile, trichostatin-A differentially affects cancer and normal (non-cancer) cells. Trichostatin-A can activate ERK1/2 to prevent TGF β 1 and serum-starvation induced apoptosis in normal cells, while potentiating apoptosis in the cancer cells [22]. It also blocks proliferation in various carcinoma cell lines [22; 23], delays tumor growth in xenograft models [24; 25], and was shown to shift the tumor phenotype from carcinoma to benign in a carcinogen-induced carcinoma model [23]. Trichostatin-A treatment prior to implantation also augmented the innate anti-tumor response in a glioblastoma xenograft model by potentiating natural killer (NK) cell-mediated lysis [24]. While the anti-tumor benefits of trichostatin-A alone appear to be modest, trichostatin-A may be more effective in combination therapy. Trichostatin-A was found to have a synergistic effect with metformin in an osteocarcinoma model [25], re-sensitized breast cancer cells to doxorubin [26], and augmented the ability of oncolytic adenoviruses to impair cell viability in ovarian cancer cells [27].



Human Mesenchymal Stem Cell (MSC) maintenance: Potential benefit (cell culture)

Trichostatin-A treatment of MSCs suppresses the morphological changes and loss of proliferation capacity that typically occurs as they are passaged in cell culture [28]. It also stabilized the expression of pluripotent genes (i.e. Oct4, Sox2, Nanog) and allowed for the maintenance of their multipotent differentiation capacity.

Kidney disease: Potential minor benefit (rodents)

Trichostatin-A helps protect renal function by preventing the loss of Klotho expression that normally accompanies kidney damage (through loss of promoter acetylation). Renal dysfunction is typically associated with the development of proteinuria/albuminuria, and these patients have lower Klotho levels [29]. Trichostatin-A can protect against albumin-mediated downregulation of Klotho in cultured renal cells, and chronic administration (0.5 mg/kg body i.p daily for 6 weeks) was reno-protective in a chronic kidney disease by preserving Klotho expression *in vivo* [30]. As with other agents that mediate protection through Klotho, trichostatin-A would only be expected to be useful in mitigating damage in early stage disease.

Oxidative stress-mediated organ damage: Potential benefit (rodents)

Trichostatin-A has been shown to protect against ischemic/oxidative stress related damage through induction of anti-oxidant and anti-inflammatory pathways.

Cardiovascular: In a coronary-artery ischemia model, trichostatin-A pretreatment reduced myocardial infarct size, and **reduced the level of oxidative stress** markers (ROS, MDA) [31]. The oxidative stress mitigation was related to the induction of SOD and FoxO3a, which was likely mediated by induction of Nrf2 and Klotho, respectively. Trichostatin-A can also suppress aberrant angiogenesis by reducing expression of Nox4 redox signaling [32].

Lung: In a lung ischemia/reperfusion injury model, trichostatin-A dose-dependently reduced vascular permeability, edema, and arterial hypertension [33]. This was associated with a reduction in neutrophil-mediated oxidative stress and inflammation.



Safety: Limited information about long-term safety in animals, and initial safety testing in humans is ongoing. Has pleiotropic effects, which may include inhibition of oligodendrocyte differentiation and altered expression of pro-atherosclerotic genes.

Types of evidence:

- Several laboratory studies

Trichostatin-A **has not yet been tested in humans**, but the first Phase I trial began recruiting patients in October 2018. Most animal studies involve acute treatment with trichostatin-A, so its **long-term safety profile is largely unknown** [34]. A single dose (1 mg/kg) in pregnant mice did not adversely affect the development of the pups [35]. Repeated dosing up to 5 mg/kg has not led to any measurable signs of toxicity in rodents [7; 23; 36]. However, due to the pleiotropic effects of HDAC inhibitors, benefits in one cell type may be offset by harm to another cell type. For example, high dose trichostatin-A has been shown to inhibit oligodendrocyte proliferation and differentiation in culture [37], which could impair myelin repair. Trichostatin-A also regulates the expression of pro-atherosclerotic genes, and could play a role in atherogenesis [38].

Sources and dosing:

Trichostatin-a is available for research use, but not human use, from a variety of commercial suppliers.

Research underway:

Vanda Pharmaceuticals is sponsoring a dose escalation Phase 1 trial for trichostatin-A (VTR-297) for patients with hematological malignancies. The first patient was randomized in October [2018](#).

Search terms:

Pubmed, Google: Trichostatin-A + neurodegeneration, dementia, Alzheimer's disease, neuroprotection, aging, lifespan, ApoE4, cancer, klotho, cardiovascular, safety, pharmacokinetics, clinical trial

Websites visited for Trichostatin-A:

- [PubChem](#)



References:

1. Dokmanovic M, Clarke C, Marks PA (2007) Histone Deacetylase Inhibitors: Overview and Perspectives. *Molecular Cancer Research* 5, 981-989. <http://mcr.aacrjournals.org/content/molcanres/5/10/981.full.pdf>
2. Fontán-Lozano Á, Romero-Granados R, Troncoso J *et al.* (2008) Histone deacetylase inhibitors improve learning consolidation in young and in KA-induced-neurodegeneration and SAMP-8-mutant mice. *Molecular and Cellular Neuroscience* 39, 193-201. <http://www.sciencedirect.com/science/article/pii/S1044743108001620>
3. Gaub P, Joshi Y, Wuttke A *et al.* (2011) The histone acetyltransferase p300 promotes intrinsic axonal regeneration. *Brain* 134, 2134-2148. <http://dx.doi.org/10.1093/brain/awr142>
4. Vadnal J, Houston S, Bhatta S *et al.* (2012) Transcriptional signatures mediated by acetylation overlap with early-stage Alzheimer's disease. *Experimental Brain Research* 221, 287-297. <https://doi.org/10.1007/s00221-012-3172-y>
5. Wang B-Y, Zhong Y, Zhao Z *et al.* (2014) Epigenetic suppression of hippocampal BDNF mediates the memory deficiency induced by amyloid fibrils. *Pharmacology Biochemistry and Behavior* 126, 83-89. <http://www.sciencedirect.com/science/article/pii/S0091305714002615>
6. Francis Y, Fà M, Ashraf H *et al.* (2009) Dysregulation of Histone Acetylation in the APP/PS1 Mouse Model of Alzheimer's Disease *Journal of Alzheimer's Disease* 18, 131-139
7. Yang W, Chauhan A, Wegiel J *et al.* (2014) Effect of Trichostatin A on Gelsolin Levels, Proteolysis of Amyloid Precursor Protein, and Amyloid Beta-Protein Load in the Brain of Transgenic Mouse Model of Alzheimer's Disease. *Current Alzheimer Research* 11, 1002-1011. <http://www.eurekaselect.com.eresources.mssm.edu/node/125925/article>
8. Suo H, Wang P, Tong J *et al.* (2015) NRSF is an essential mediator for the neuroprotection of trichostatin A in the MPTP mouse model of Parkinson's disease. *Neuropharmacology* 99, 67-78. <http://www.sciencedirect.com/science/article/pii/S0028390815300216>
9. Belakavadi M, Dell J, Grover GJ *et al.* (2011) Thyroid hormone suppression of β -amyloid precursor protein gene expression in the brain involves multiple epigenetic regulatory events. *Molecular and Cellular Endocrinology* 339, 72-80. <http://www.sciencedirect.com/science/article/pii/S0303720711001766>
10. Zhu M, Li W-W, Lu C-Z (2014) Histone Decacetylase Inhibitors Prevent Mitochondrial Fragmentation and Elicit Early Neuroprotection against MPP+. *CNS Neuroscience & Therapeutics* 20, 308-316. <https://onlinelibrary.wiley.com/doi/abs/10.1111/cns.12217>
11. Wang Y, Wang X, Liu L *et al.* (2009) HDAC inhibitor trichostatin A-inhibited survival of dopaminergic neuronal cells. *Neuroscience Letters* 467, 212-216. <http://www.sciencedirect.com/science/article/pii/S0304394009013573>
12. Giral A, Puigdel·l·ivol M, Carret·n O *et al.* (2012) Long-term memory deficits in Huntington's disease are associated with reduced CBP histone acetylase activity. *Human Molecular Genetics* 21, 1203-1216. <http://dx.doi.org/10.1093/hmg/ddr552>
13. Dompierre JP, Godin JD, Charrin BC *et al.* (2007) Histone Deacetylase 6 Inhibition Compensates for the Transport Deficit in Huntington's Disease by Increasing Tubulin Acetylation. *The Journal of Neuroscience* 27, 3571-3583. <http://www.jneurosci.org/content/jneuro/27/13/3571.full.pdf>



14. Ma X-H, Gao Q, Jia Z *et al.* (2015) Neuroprotective capabilities of TSA against cerebral ischemia/reperfusion injury via PI3K/Akt signaling pathway in rats. *International Journal of Neuroscience* 125, 140-146. <https://doi.org/10.3109/00207454.2014.912217>
15. Wang B, Zhu X, Kim Y *et al.* (2012) Histone deacetylase inhibition activates transcription factor Nrf2 and protects against cerebral ischemic damage. *Free radical biology & medicine* 52, 928-936. <https://www.ncbi.nlm.nih.gov/pubmed/22226832>
<https://www.ncbi.nlm.nih.gov/pmc/PMC6010182/>
16. Das U, Scott DA, Ganguly A *et al.* (2013) Activity-induced convergence of APP and BACE-1 in acidic microdomains via an endocytosis-dependent pathway. *Neuron* 79, 447-460. <https://www.ncbi.nlm.nih.gov/pubmed/23931995>
<https://www.ncbi.nlm.nih.gov/pmc/PMC3741682/>
17. Prasad H, Rao R (2018) Amyloid clearance defect in ApoE4 astrocytes is reversed by epigenetic correction of endosomal pH. *Proceedings of the National Academy of Sciences* 115, E6640-E6649. <http://www.pnas.org/content/pnas/115/28/E6640.full.pdf>
18. Calvert S, Tacutu R, Sharifi S *et al.* (2016) A network pharmacology approach reveals new candidate caloric restriction mimetics in *C. elegans*. *Aging Cell* 15, 256-266. <https://onlinelibrary.wiley.com/doi/abs/10.1111/ace1.12432>
19. Tao D, Lu J, Sun H *et al.* (2004) Trichostatin A Extends the Lifespan of *Drosophila melanogaster* by Elevating hsp22 Expression. *Acta Biochimica et Biophysica Sinica* 36, 618-622. <http://dx.doi.org/10.1093/abbs/36.9.618>
20. Zhao Y, Sun H, Lu J *et al.* (2005) Lifespan extension and elevated hsp gene expression in *Drosophila* caused by histone deacetylase inhibitors. *Journal of Experimental Biology* 208, 697-705. <http://jeb.biologists.org/content/jexbio/208/4/697.full.pdf>
21. Prasad H, Rao R (2018) Histone deacetylase-mediated regulation of endolysosomal pH. *Journal of Biological Chemistry* 293, 6721-6735. <http://www.jbc.org/content/293/18/6721.abstract>
22. Zhang Y, Yu G, Wang D *et al.* (2011) ERK1/2 activation plays important roles in the opposite effects of Trichostatin A in non-cancer and cancer cells. *Toxicol* 57, 932-937. <http://www.sciencedirect.com/science/article/pii/S0041010111000948>
23. Vigushin DM, Ali S, Pace PE *et al.* (2001) Trichostatin A Is a Histone Deacetylase Inhibitor with Potent Antitumor Activity against Breast Cancer in Vivo. *Clinical Cancer Research* 7, 971-976. <http://clincancerres.aacrjournals.org/content/clincanres/7/4/971.full.pdf>
24. Horing E, Podlech O, Silkenstedt B *et al.* (2013) The Histone Deacetylase Inhibitor Trichostatin A Promotes Apoptosis and Antitumor Immunity in Glioblastoma Cells. *Anticancer Research* 33, 1351-1360. <http://ar.iijournals.org/content/33/4/1351.abstract>
25. Duo J, Ma Y, Wang G *et al.* (2013) Metformin Synergistically Enhances Antitumor Activity of Histone Deacetylase Inhibitor Trichostatin A Against Osteosarcoma Cell Line. *DNA and Cell Biology* 32, 156-164. <https://www.liebertpub.com/doi/abs/10.1089/dna.2012.1926>
26. Ponnusamy L, Mahalingaiah PKS, Chang Y-W *et al.* (2018) Reversal of epigenetic aberrations associated with the acquisition of doxorubicin resistance restores drug sensitivity in breast cancer cells. *European Journal of Pharmaceutical Sciences* 123, 56-69. <http://www.sciencedirect.com/science/article/pii/S0928098718303269>



27. Hulin-Curtis SL, Davies JA, Jones R *et al.* (2018) Histone deacetylase inhibitor trichostatin A sensitises cisplatin-resistant ovarian cancer cells to oncolytic adenovirus. *Oncotarget* 9, 26328-26341. <https://www.ncbi.nlm.nih.gov/pubmed/29899862> <https://www.ncbi.nlm.nih.gov/pmc/PMC5995174/>
28. Han B, Li J, Li Z *et al.* (2013) Trichostatin A stabilizes the expression of pluripotent genes in human mesenchymal stem cells during ex vivo expansion. *PLoS one* 8, e81781-e81781. <https://www.ncbi.nlm.nih.gov/pubmed/24312356> <https://www.ncbi.nlm.nih.gov/pmc/PMC3842316/>
29. Fernandez-Fernandez B, Izquierdo MC, Valiño-Rivas L *et al.* (2018) Albumin downregulates Klotho in tubular cells. *Nephrology Dialysis Transplantation* 33, 1712-1722. <http://dx.doi.org/10.1093/ndt/gfx376>
30. Lin W, Li Y, chen F *et al.* (2017) Klotho preservation via histone deacetylase inhibition attenuates chronic kidney disease-associated bone injury in mice. *Scientific Reports* 7, 46195. <https://doi.org/10.1038/srep46195>
31. Guo Y, Li Z, Shi C *et al.* (2017) Trichostatin A attenuates oxidative stress-mediated myocardial injury through the FoxO3a signaling pathway. *International journal of molecular medicine* 40, 999-1008. <https://www.ncbi.nlm.nih.gov/pubmed/28849190> <https://www.ncbi.nlm.nih.gov/pmc/PMC5593460/>
32. Hakami NY, Dusting GJ, Peshavariya HM (2016) Trichostatin A, a histone deacetylase inhibitor suppresses NADPH Oxidase 4-Derived Redox Signalling and Angiogenesis. *Journal of cellular and molecular medicine* 20, 1932-1944. <https://www.ncbi.nlm.nih.gov/pubmed/27297729> <https://www.ncbi.nlm.nih.gov/pmc/PMC5020625/>
33. Hsu H-H, Wu S-Y, Tang S-E *et al.* (2015) Protection against reperfusion lung injury via abrogating multiple signaling cascades by trichostatin A. *International Immunopharmacology* 25, 267-275. <http://www.sciencedirect.com/science/article/pii/S1567576915000600>
34. Sanderson L, Taylor GW, Aboagye EO *et al.* (2004) Plasma pharmacokinetics and metabolism of the histone deacetylase inhibitor trichostatin-a after intraperitoneal administration to mice. *Drug Metabolism and Disposition* 32, 1132-1138. <http://dmd.aspetjournals.org/content/dmd/32/10/1132.full.pdf>
35. Nervi C, Borello U, Fazi F *et al.* (2001) Inhibition of Histone Deacetylase Activity by Trichostatin A Modulates Gene Expression during Mouse Embryogenesis without Apparent Toxicity. *Cancer Research* 61, 1247-1249. <http://cancerres.aacrjournals.org/content/canres/61/4/1247.full.pdf>
36. Sharma S, Taliyan R, Ramagiri S (2015) Histone Deacetylase Inhibitor, Trichostatin A, Improves Learning and Memory in High-Fat Diet-Induced Cognitive Deficits in Mice. *Journal of Molecular Neuroscience* 56, 1-11. <https://doi.org/10.1007/s12031-014-0461-x>
37. Conway GD, O'Bara MA, Vedia BH *et al.* (2012) Histone deacetylase activity is required for human oligodendrocyte progenitor differentiation. *Glia* 60, 1944-1953. <https://onlinelibrary.wiley.com/doi/abs/10.1002/glia.22410>
38. Choi J-H, Nam K-H, Kim J *et al.* (2005) Trichostatin A Exacerbates Atherosclerosis in Low Density Lipoprotein Receptor-Deficient Mice. *Arteriosclerosis, Thrombosis, and Vascular Biology* 25, 2404-2409. <https://doi.org/10.1161/01.ATV.0000184758.07257.88>



Disclaimer: Cognitive Vitality Reports® do not provide, and should not be used for, medical advice, diagnosis, or treatment. You should consult with your healthcare providers when making decisions regarding your health. Your use of these reports constitutes your agreement to the [Terms & Conditions](#).

If you have suggestions for drugs, drugs-in-development, supplements, nutraceuticals, or food/drink with neuroprotective properties that warrant in-depth reviews by ADDF's Aging and Alzheimer's Prevention Program, please contact INFO@alzdiscovery.org. To view our official ratings, visit [Cognitive Vitality's Rating page](#).